

Deconvoluting AMP-activated protein kinase (AMPK) adenine nucleotide binding and sensing

Received for publication, April 25, 2017, and in revised form, May 31, 2017. Published, Papers in Press, June 14, 2017, DOI 10.1074/jbc.M117.793018

Xin Gu[‡], Yan Yan^{‡§}, Scott J. Novick[¶], Amanda Kovach[‡], Devrishi Goswami[¶], Jiyuan Ke[‡], M. H. Eileen Tan^{‡||}, Lili Wang[‡], Xiaodan Li[‡], Parker W. de Waal[‡], Martin R. Webb^{||}, Patrick R. Griffin[¶], H. Eric Xu^{‡§}, and Karsten Melcher^{‡1}

From the [‡]Laboratories of Structural Sciences and Structural Biology and Biochemistry, Center for Cancer and Cell Biology, Van Andel Research Institute, Grand Rapids, Michigan 49503, the [§]VARI-SIMM Center, Center for Structure and Function of Drug Targets, The CAS Key Laboratory of Receptor Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences (CAS), Shanghai 201203, China, the [¶]Department of Molecular Medicine, Translational Research Institute, The Scripps Research Institute, Jupiter, Florida 33458, and ^{||}The Francis Crick Institute, 1 Midland Road, London NW1 1AT, United Kingdom

Edited by Wolfgang Peti

AMP-activated protein kinase (AMPK) is a central cellular energy sensor that adapts metabolism and growth to the energy state of the cell. AMPK senses the ratio of adenine nucleotides (adenylate energy charge) by competitive binding of AMP, ADP, and ATP to three sites (CBS1, CBS3, and CBS4) in its γ -subunit. Because these three binding sites are functionally interconnected, it remains unclear how nucleotides bind to individual sites, which nucleotides occupy each site under physiological conditions, and how binding to one site affects binding to the other sites. Here, we comprehensively analyze nucleotide binding to wild-type and mutant AMPK protein complexes by quantitative competition assays and by hydrogen-deuterium exchange MS. We also demonstrate that NADPH, in addition to the known AMPK ligand NADH, directly and competitively binds AMPK at the AMP-sensing CBS3 site. Our findings reveal how AMP binding to one site affects the conformation and adenine nucleotide binding at the other two sites and establish CBS3, and not CBS1, as the high affinity exchangeable AMP/ADP/ATP-binding site. We further show that AMP binding at CBS4 increases AMP binding at CBS3 by 2 orders of magnitude and reverses the AMP/ATP preference of CBS3. Together, these results illustrate how the three CBS sites collaborate to enable highly sensitive detection of cellular energy states to maintain the tight ATP homeostasis required for cellular metabolism.

This work was supported by the Francis Crick Institute with funding from Cancer Research UK Grant FC001211, United Kingdom Medical Research Council Grant FC001211, Wellcome Trust Grant FC001211 (to M. R. W.), grants from the Van Andel Research Institute (to H. E. X. and K. M.), Ministry of Science and Technology (China) Grants 2012ZX09301001, 2012CB910403, 2013CB910600, XDB08020303, and 2013ZX09507001, National Science Foundation Grant 91217311 (to H. E. X.), and National Institute of Health Grants DK071662 (to H. E. X.) and GM102545 and GM104212 (to K. M.). The authors declare that they have no conflicts of interest with the contents of this article. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Author's Choice—Final version free via Creative Commons CC-BY license. This article contains supplemental Tables S1–S3.

¹ To whom correspondence should be addressed: Laboratory of Structural Biology and Biochemistry, Center of Cancer and Cell Biology, Van Andel Research Institute, 333 Bostwick Ave. Northeast, Grand Rapids, MI 49503. Tel.: 616-234-5699; Fax: 616-234-5170; Email: karsten.melcher@vai.org.

ATP homeostasis is critically important for cellular functions and is maintained at a tight concentration range by the regulatory protein kinase AMP-activated protein kinase (AMPK),² a therapeutic target for metabolic diseases and cancer (1–7). Because of this tight range, AMPK senses the energy state of the cell largely as ratio of AMP to ATP, which changes much more dramatically than ATP levels under energy stress, whereas ADP levels change relatively more modestly. High levels of AMP activate AMPK by three different mechanisms: direct allosteric activation of its kinase activity (8, 9), increasing activation loop phosphorylation by upstream kinase (10, 11), and protection of the phosphorylated activation loop against dephosphorylation (12, 13). High levels of ADP also protect against activation loop dephosphorylation (although 5–10 times less potently than AMP for the γ 1 and γ 3 isoforms) and do not stimulate direct kinase activation as AMP (8, 14–17).

AMPK consists of three subunits, a kinase domain-containing α -subunit, a carbohydrate-binding module (CBM)-containing β -subunit, and an adenine nucleotide-binding γ -subunit. The γ -subunit contains four cystathionine β -synthetase (CBS) motifs, which function in pairs known as Bateman domains as universal adenine nucleotide-binding sites (18). CBS1 and CBS2 form Bateman domain 1, whereas CBS3 and CBS4 form Bateman domain 2 (note that CBS1, -2, -3, and -4 were previously (19) known as sites 2, 4, 1, and 3). Crystal structures and binding assays have established that CBS2, in which the conserved, ribose-binding aspartate is mutated to arginine, fails to bind adenine nucleotides (15, 19–21). On the other hand, AMP is tightly bound to CBS4 (19, 20) as wild-type AMPK and *cbs1* and *cbs3* mutant AMPK co-purify with an equimolar amount of AMP, whereas a *cbs4* mutant only purifies with trace amounts of nucleotides (20). Therefore CBS4 likely does not exchange AMP against ATP in the presence of physiological AMP levels (19). However, under artificial condi-

² The abbreviations used are: AMPK, AMP-activated protein kinase; α RIM, α -regulatory subunit interaction motif; AXP, AMP/ADP/ATP adenine nucleotides; CBM, carbohydrate-binding module; CBS, cystathionine β -synthetase; deac-ADP, 3'-(7-diethylaminocoumarin-3-carbonylamino)-3'-deoxyadenosine-5'-diphosphate; HDX-MS, hydrogen deuterium exchange mass spectrometry; ITC, isothermal titration calorimetry; MBP, maltose-binding protein; PDB, Protein Data Bank.

tions, *i.e.* the complete absence of AMP, prebound AMP at CBS4 can very slowly exchange against ATP in solution ($\sim 40\%$ exchange after removal of free nucleotides, followed by 6 h incubation with 5 mM ATP) (20). The remaining two CBS sites (CBS1 and CBS3) are thought to be exchangeable as biochemical binding and competition experiments confirmed the existence of two readily exchangeable adenine nucleotide (AXP)-binding sites in purified recombinant AMPK (15, 18, 19, 22). Mutations in key AXP-binding residues in the $\gamma 2$ - and $\gamma 3$ -AMPK isoforms increase basal AMPK activity and are causally associated with glycogen storage diseases and Wolf-Parkinson-White cardiomyopathy (23–30).

AMP-occupied CBS3 is directly bound by a loop (α RIM) adjacent to the autoinhibitory domain of the α -subunit (15, 31), suggesting that the α RIM senses AMP at CBS3. Consistently, mutating key α RIM residues compromises AMPK activation (31). Moreover, AMP increases and ATP decreases the interaction between the isolated α RIM loop and core-AMPK (γ -subunit plus scaffolding C termini of the α - and β -subunit), and AMP stimulation of the interaction requires the intact α RIM (32). In addition to mutations in CBS3, also mutations in CBS4, which does not interact with the α RIM or other parts of the α - or β -subunits, abolish allosteric kinase activation by AMP (20), yet the molecular basis for the CBS4 requirement has remained elusive. When ATP was soaked into crystals of AMP-bound core-AMPK, ATP occupied CBS1 and CBS3, whereas CBS4 remained AMP bound (19, 20). In contrast, when AMP-depleted core-AMPK was co-crystallized with ATP, ATP bound to CBS1 and CBS4, CBS3 was unoccupied, and the γ -subunit underwent a noticeable conformational change (20).

One of the two exchangeable AXP-binding sites binds both AMP and Mg^{2+} -free ATP with micromolar affinity ("high affinity site"), whereas the other site binds AXP much more weakly (hundreds of micromolar (15, 19, 22)), and likely with positive cooperativity (18). Recent surface plasmon resonance (Biacore) data have revealed that ATP binds AMPK in the presence of Mg^{2+} , which shields charge interactions of ATP phosphates with CBS residues, with single site kinetics and a binding constant in the 100 μM range (22). This low affinity of Mg^{2+} -ATP would be consistent with preferential AMP-binding at the high affinity binding site to allow AMP exchange at nucleotide concentrations that reflect cellular conditions of energy excess and energy stress (14). Currently, it is poorly understood which affinity state maps to which CBS site, how nucleotide binding at one site affects binding at the other site(s), and what adenine nucleotide binds to each of the three functional CBS sites at physiological concentrations of ATP, ADP, and AMP. Focusing on AMP and ATP, we combined fluorescence binding and competition experiments of wild-type and mutant AMPK with hydrogen deuterium exchange mass spectrometry (HDX-MS). These experiments allowed us to (i) identify CBS3 as a high affinity exchangeable AXP-binding site as well as binding site for NADPH, (ii) map the CBS conformational connectivity and demonstrate that preferential AMP-binding at CBS3 requires AMP-bound CBS4, and (iii) suggest that under physiological conditions only CBS3 significantly exchanges between AMP and ATP.

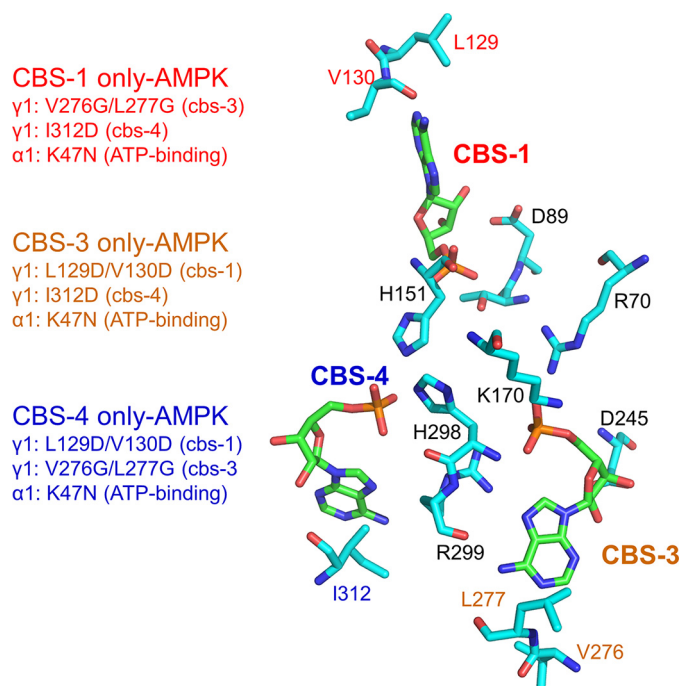


Figure 1. Binding of AMP to the $\gamma 1$ subunit. Stick presentation of the three AMP molecules (green carbon atoms) and key AMP-binding residues of AMPK (cyan carbon atoms). Oxygen is colored in red, nitrogen in blue, and phosphorus in orange. The phosphate groups of the 3 AMP molecules are coordinately bound by a set of charged amino acids, whereas the ribose and adenine rings face away from each other and are bound by CBS-specific residues. Mutations of the indicated adenine-binding residues therefore selectively block AMP binding to individual CBS sites. Note that the adenine-binding mutations at CBS3 and CBS4 are each sufficient, even at the high concentration of adenine nucleotides in cells, to completely abolish AMPK activation by AMP without affecting AMPK catalytic activity in the absence of AMP (18).

Results

Isolated CBS motifs bind AXP weakly and with altered AMP/ATP specificity

The phosphate groups of the three AMP molecules in AMP-bound AMPK face toward each other and are coordinately bound by a set of charged amino acids, whereas the adenine rings face away from each other and are bound by CBS-specific residues (Fig. 1). Specifically, His¹⁵¹ binds the phosphate groups of both AMP at CBS1 and CBS4, whereas His²⁹⁸ forms hydrogen bonds with the phosphates at both CBS3 and CBS4, suggesting that binding to one CBS motif could affect binding to the other sites. To characterize nucleotide binding at individual CBS motifs in the context of holo-AMPK, but absence of other nucleotide-binding sites, we generated a set of three AMPK variants in which we mutated the ATP-binding lysine in the kinase domain (α_1 -K47N) plus adenine-binding residues previously shown to be required for nucleotide binding (20) at two of three CBS sites, thus only one functional CBS remains (Fig. 1). All mutant proteins were stable and could be purified as stoichiometric complexes from size exclusion chromatography columns (Fig. 2). Similar to a previous approach (19), we determined relative nucleotide affinities by incubating wild-type and mutant proteins with a fluorescently labeled adenine nucleotide analog and competed the interaction with unlabeled AMP or ATP. Incubation of 500 nM 3'-(7-diethylaminocoumarin-3-

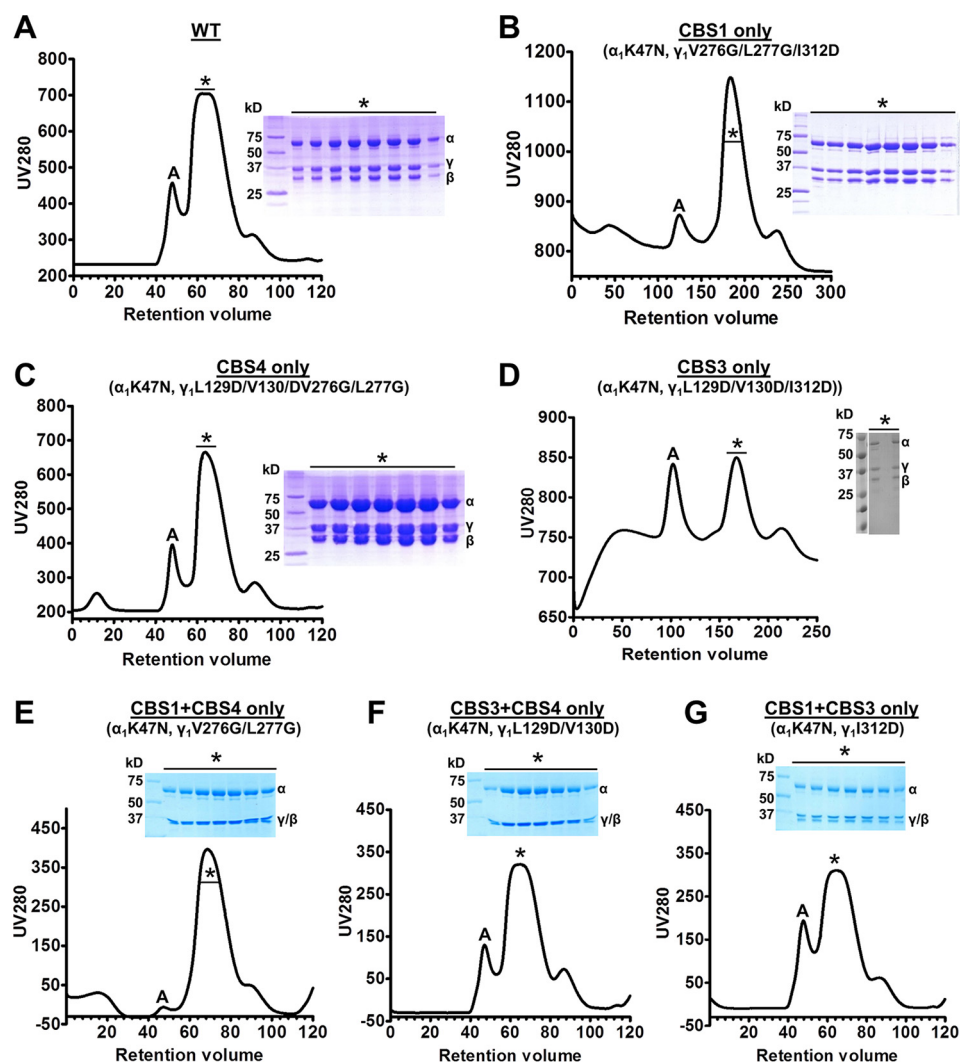


Figure 2. Size exclusion chromatograms and SDS-PAGE of AMPK mutant proteins. A, C, and E–G, 120-ml column; B and D, 300-ml column; *, monomeric $\alpha_1\beta_2\gamma_1$ fractions; A, aggregate/oligomers.

carbonylamin)-3'-deoxyadenosine-5'-diphosphate (deac-ADP; see "Experimental procedures") (33) with 4 μM of the recombinant human $\alpha_1\beta_2\gamma_1$ -isoform of AMPK resulted in a strong increase in deac-ADP fluorescence and a shift of the emission maximum to a shorter wavelength (Fig. 3A), indicating that deac-ADP binds wild-type and the three mutant AMPK proteins.

As controls, we first incubated deac-ADP with AMPK in which the adenine-binding residues at all three CBS sites were mutated (plus K47N in the kinase domain), which resulted in a small increase in fluorescence (Fig. 3B). This indicated that deac-ADP, in which the ADP-ribose is modified by a bulky coumarin derivative, has residual AMPK-binding activity. This binding signal could not be competed with a 10,000-fold molar excess of AMP, consistent with the reported inability of AMP to bind to this triple cbs mutant (cbs1, L129D/V130D; cbs2, V276G/L277G; cbs3, I312D) (20). Neither could ATP compete with the increase in fluorescence, but surprisingly caused a minor increase in fluorescence that was diminished by introduction of an additional cbs4 mutation, S314A (Fig. 3C). Second, in excellent agreement with previous studies (15, 19, 22),

Mg^{2+} -AMP bound wild-type AMPK, and competed the binding of deac-ADP to wild-type AMPK, with biphasic kinetics, consistent with two exchangeable binding sites, with K_d values of 2.1 μM (high affinity site) and $\sim 290 \mu\text{M}$ (low affinity site) (Fig. 4, A–C), whereas Mg^{2+} -ATP competed with monophasic kinetics, consistent with similar affinities for both exchangeable sites (Fig. 4B).

We then tested the ability of AMP and ATP to compete deac-ADP from the AMPK mutant proteins. In contrast to wild-type AMPK, the "non-exchangeable" AMP-binding site CBS4 became exchangeable in the cbs1/cbs3 mutant, as it was efficiently bound by deac-ADP, and binding could be competed with IC_{50} values of 3.8 μM (AMP) and 14.8 μM (ATP) in the absence of Mg^{2+} , and 1.9 μM for AMP in the presence of Mg^{2+} (Fig. 5A; note that Mg^{2+} -ATP inefficiently competed deac-ADP from the CBS4 only mutant, resulting in an unreliable IC_{50} value for this measurement). Both AMPK cbs3/cbs4 (CBS1 only) and AMPK cbs1/cbs4 (CBS3 only) bound ATP with low to moderate affinity and AMP with very low affinity. Especially CBS3 in the absence of CBS4 and CBS1 bound AMP with an IC_{50} value that is well below physiological AMP concentrations

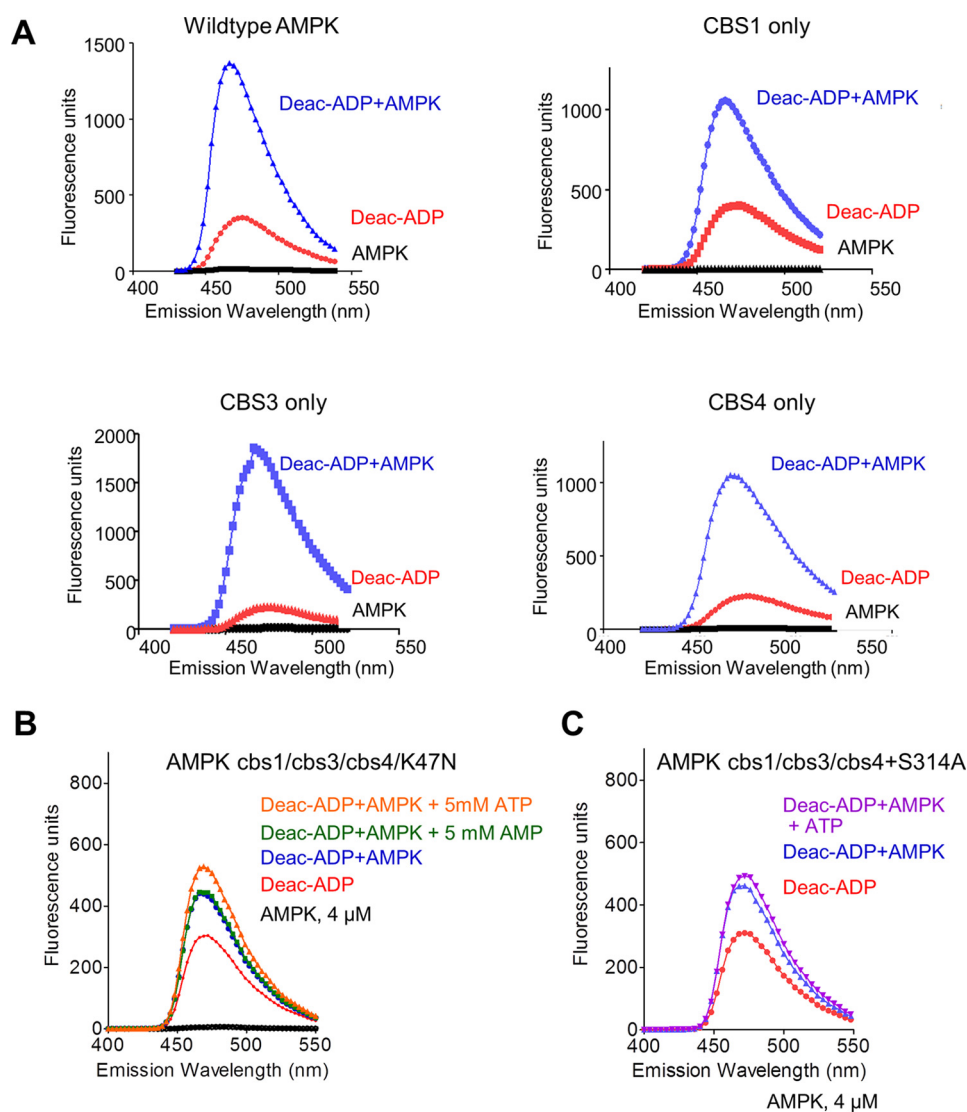


Figure 3. Fluorescence spectra of wild-type and mutant AMPK/deac-ADP and competition by AMP and ATP. A, emission spectra of 0.5 μM deac-ADP in the absence and presence of wild-type and mutant AMPK proteins in which only one of the CBS sites remained functional (CBS1 only: $\alpha 1$ -K47N, $\gamma 1$ -V276G/L277G/I312D; CBS3 only: $\alpha 1$ -L129D/V130D/I312D; CBS4 only: $\alpha 1$ -K47N, $\gamma 1$ -L129D/V130D/V276G/L277G). Excitation: 430 nm. B and C, emission spectra of 0.5 μM deac-ADP in the absence and presence of 4 μM $\alpha 1\beta 2\gamma 1$ AMPK ($\alpha 1$ -K47N, $\gamma 1$ -L129D/V130D/V276G/L277G/I312D) (B) or $\alpha 1\beta 2\gamma 1$ AMPK ($\alpha 1$ -K47N, $\gamma 1$ -L129D/V130D/V276G/L277G/I312D/S314A) (C) and 5 mM AMP or ATP.

and with a more than 8-fold preference of ATP over AMP in the presence of Mg^{2+} (and more than 90-fold in its absence). Given that the physiological concentration of ATP is close to 2 orders of magnitude higher than that of AMP, CBS3 would be constitutively bound by ATP in the absence of functional CBS4 and CBS1.

Next we determined binding of AMP and ATP to AMPK in which only the ATP-binding site in the kinase domain and a single CBS site were mutated, leaving intact two binding sites in all three combinations. Importantly, Chen *et al.* (20) had shown that in these mutants CBS4 remains tightly AMP-bound during protein purification. In agreement with this report, when we left CBS3 and CBS4 intact ($\alpha 1$ -K47N, $\gamma 1$ -L129D/V130D), AMP and ATP competed a single site (*i.e.* data cannot be modeled to two sites), confirming that the presence of wild-type CBS3 is sufficient to allow CBS4 to bind AMP tightly enough to not noticeably exchange during competition. In the presence of wild-type CBS4, CBS3 bound AMP with relatively high affinity

(16.9 μM) and ATP with much lower affinity (153 μM) (Fig. 5B), resembling the high affinity exchangeable binding site in the context of wild-type AMPK. In contrast, when we mutated the catalytic site plus CBS3 (CBS1 + CBS4 intact), the competition again fit only a single site model, yet the affinity for AMP remained weak ($\text{IC}_{50} = 160 \mu\text{M}$) and below that of ATP ($\text{IC}_{50} = 78 \mu\text{M}$) (Fig. 5B), resembling more closely the low affinity exchangeable AXP-binding site. Finally, when only CBS1 and CBS3 remained functional, we could fit the competition data only as a single site with low affinity for ATP and very low affinity for AMP (Fig. 5B), similar to what we have found for the exchangeable low affinity binding site in wild-type AMPK. Collectively, these data indicate that (i) either a functional CBS1 or a functional CBS3 is sufficient to allow CBS4 to bind AMP non-reversibly during competition, that (ii) in the presence of functional CBS4, CBS3 resembles more the high affinity and CBS1 the low affinity exchangeable site, and (iii) that in the presence of only CBS1 and CBS3 neither formed a high affinity binding

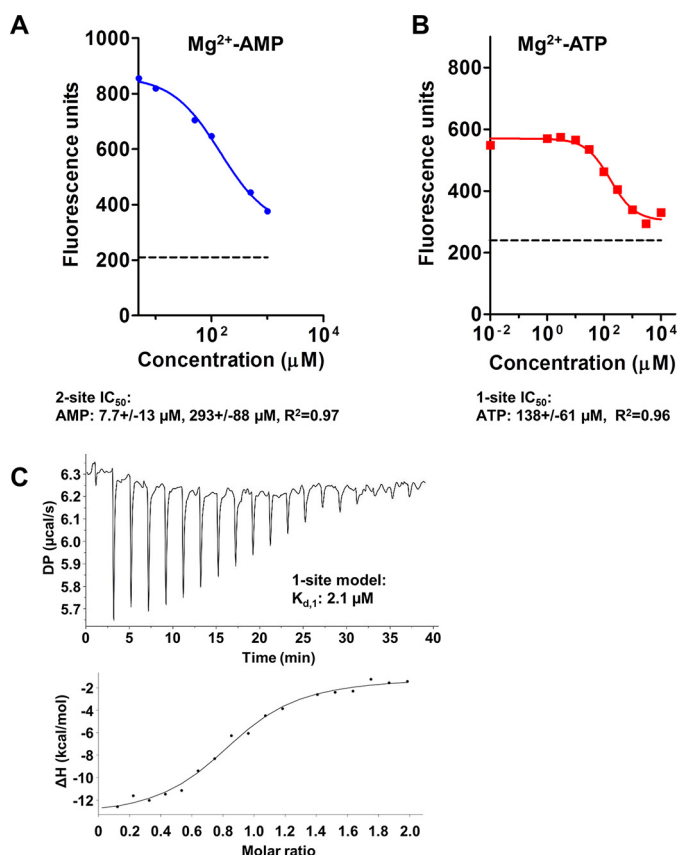


Figure 4. Nucleotide binding to wild-type AMPK. A and B, competition of $0.5 \mu\text{M}$ deac-ADP bound to $\alpha 1\beta 2\gamma 1$ AMPK by unlabeled Mg^{2+} -AMP (A) and Mg^{2+} -ATP (B). The dashed line indicates the fluorescence signal of unbound deac-ADP. Because fluorescence competition failed to unambiguously determine the IC_{50} value for the high affinity AMP-binding site, we independently determined its affinity by ITC (C).

site, but both most likely formed low affinity binding sites that could not be computationally deconvoluted.

AXP binding to a single CBS motif modulates the conformation of all three CBS sites as well as the CBM

To directly test conformational stabilization, we analyzed wild-type and mutant AMPK by HDX-MS. HDX-MS measures the backbone amide exchange of hydrogen against deuterium in deuterated buffers. Ligand binding typically protects binding sites against deuterium exchange and can also induce conformational changes that alter protection patterns. Incubation of wild-type AMPK with 2.5 mM AMP dramatically reduced hydrogen/deuterium exchange at CBS3 and CBS4, strongly at CBS1, and moderately at the catalytic center of the kinase domain (catalytic loop, activation loop, and parts of helices αB , αD , and αF ; Fig. 6A and supplemental Tables S1–S3). Because AMP at CBS4 exchanges very slowly, the strong decrease in HDX at CBS4 is likely predominantly or completely due to its conformational stabilization upon AMP binding to CBS1 and CBS3.

Unexpectedly, the CBM of the β -subunit was also strongly protected (Fig. 6A, left panel, and supplemental Table S3), indicating that the adenine nucleotide-binding sites in the γ -subunit and the CBM are conformationally connected, even though these domains are located on opposite sides of the

AMPK heterotrimer and do not contact each other. Conversely, when we incubated AMPK with the Merck compound 991, which binds a pocket formed by the CBM and kinase domain (allosteric drug and metabolite-binding site) (34), the 991-binding site and the regulatory αC -helix became strongly stabilized, whereas the distal CBS3 in the γ -subunit became mildly destabilized (Fig. 6B). AMP-dependent protection against dephosphorylation of $p\text{-Thr}^{174}$ in the activation loop likely involves occluding $p\text{-Thr}^{174}$ by the CBM linker (15). The observed CBM stabilization by AMP binding may therefore play a direct role in this critical AMP response. When we incubated AMPK with ATP, the overall protection was similar to that with AMP. However, protection of CBS3, CBS4, and the CBM was weaker, whereas protection of CBS1 and the ATP-binding catalytic site in the kinase domain increased (Fig. 6A, right panel, and supplemental Tables S1–S3).

When we incubated the AMPK mutant protein in which CBS4 is the only functional AXP-binding site (K47N/cbs1/cbs3) with AMP, CBS4 became strongly protected, confirming that CBS4 is indeed exchangeable in the absence of CBS1 and CBS3. In addition, CBS3 became protected, indicating that AMP-binding at CBS4 strongly conformationally stabilizes CBS3 (Fig. 6C), in excellent agreement with the fluorescence competition data. In contrast to CBS3, AMP-binding at CBS4 modulated accessibility at CBS1 more mildly (Fig. 6C; note that deuterium exchange both increases and weakly decreases in different CBS1 peptides), consistent with the relatively small effect of CBS4 on CBS1 AMP-binding. A similar, but milder protection of CBS3 and CBS4 occurred for the CBS3-only (K47N/cbs1/cbs4) AMPK mutant in the presence of AMP (Fig. 6D), whereas AMP-binding to the CBS1-only (K47N/cbs3/cbs4) AMPK mutant stabilized all three binding sites (Fig. 6E).

CBS occupancy in mixtures of adenine nucleotides

All current binding and structural studies have been performed either in the presence of high concentrations of AMP or high concentrations of ATP, whereas in cells AMPK is exposed to mixtures of AMP, ADP, and ATP. An important, but unresolved, question is therefore which CBS motifs are occupied by AMP, ADP, or ATP under physiological conditions, with AMP always present, but at levels that are 10–100-fold lower than ATP levels. To probe for occupancy under physiological conditions, we incubated AMPK with two different AXP mixtures: 4.5 mM ATP, 0.4 mM ADP, 0.04 mM AMP and 3.8 mM ATP, 1.0 mM ADP, 0.3 mM AMP, which have been reported to mimic normal cellular (non-stressed) and energy stress conditions in cells, respectively (14). Consistently, the AXP ratio mimicking energy stress allosterically activated recombinantly purified AMPK 2-fold relative to the mixture mimicking energy excess (Fig. 7A), the same fold direct activation as for the standard switch from AMPK, $100 \mu\text{M}$ ATP to AMPK, $100 \mu\text{M}$ ATP + $200 \mu\text{M}$ AMP. In striking contrast to the different HDX profiles of AMPK, 2.5 mM AMP versus AMPK, 2.5 mM ATP (Fig. 6, A and B, and supplemental Tables S1–S3), the profiles in the presence of the stress and non-stress AXP mixtures were quite similar to each other and showed significant differences only in two small areas (Fig. 7B), clearly indicating that these conditions induce much more limited nucleotide exchange. Whereas HDX-MS

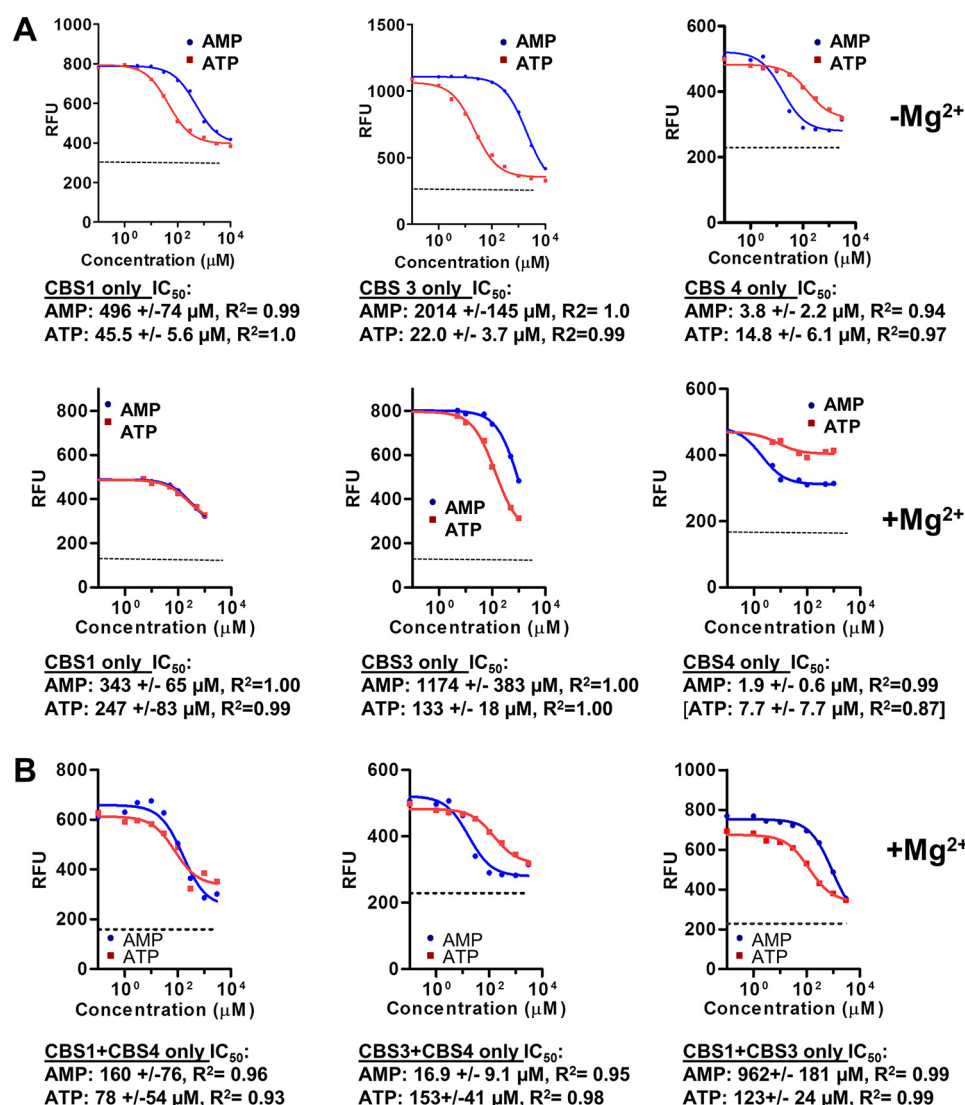


Figure 5. Nucleotide binding to individual CBSs and pairs of CBSs. Competition of 0.5 μM deac-ADP bound to 4 μM α1β2γ1 AMPK by unlabeled AMP and ATP. *A*, AMPK mutants with only a single functional adenine nucleotide-binding site (see Fig. 1). Note that in these experiments singly and doubly mutated CBS4 (I312D and I312D/S314A) behaved identically. The ability of Mg²⁺-AMP to compete deac-ADP from 4 μM AMPK with an IC₅₀ of 1.9 μM indicates that AMP still binds CBS4 with submicromolar affinity, but no longer non-exchangeably. *B*, AMPK mutants with two functional adenine nucleotide-binding sites. Samples were incubated for 30 min, excited at 430 nm, and emission recorded at 470 nm. The dashed line indicates the fluorescence signal of unbound (*i.e.* completely competed) deac-ADP.

has the potential to distinguish AMP *versus* ATP occupancy due to the unique β- and γ-phosphate groups of ATP, none of the resolved β- and γ-phosphate contacting peptides makes exclusive interactions with these groups (see Fig. 8 and [supplemental Table S1](#)) and decreases in HDX can be due to both direct nucleotide binding and to conformational stabilizations. Because our fluorescence competition data are most consistent with CBS1 (binds the more abundant ATP with equal or higher affinity than AMP) being bound by ATP under both normal and energy stress conditions, and CBS4 being essentially non-exchangeably bound to AMP, only CBS3 may undergo noticeable ATP/AMP exchange at physiological levels.

NADPH binds AMPK at the high affinity exchangeable AXP-binding site

We found that in addition to the known AMPK ligand NADH (15), NADPH can also bind AMPK. When we incubated 5 μM NADPH with 10 μM maltose-binding protein (MBP)-

tagged AMPK, NADPH fluorescence increased and shifted its emission maximum, indicative of direct binding (Fig. 9A). In contrast, incubation of NADPH with 10 μM of the MBP control did not alter the NADPH emission spectrum (Fig. 9A). To estimate the affinity for NADPH and NADH, we incubated 5 μM NADPH or 5 μM NADH with increasing concentrations of MBP-AMPK. Although the half-maximal increase of the NADPH fluorescence signal indicated a *K_d* of ≤20 μM, an increase of NADH fluorescence did not approach saturation at the highest AMPK concentration (100 μM), indicating that NADH binds AMPK only with low affinity (>>20 μM; Fig. 9B). Because NADPH is an adenine dinucleotide, we speculated that it would bind to one or more of the AXP-binding sites in the γ-subunit. Indeed 10 μM AMP almost completely competed 5 μM NADPH bound to 10 μM AMPK, indicating that NADPH binds to the high affinity exchangeable AMP-binding site (Fig. 9C). Conversely, ATP was also able to compete NADPH, but only at much higher concentrations (Fig. 9D). We therefore

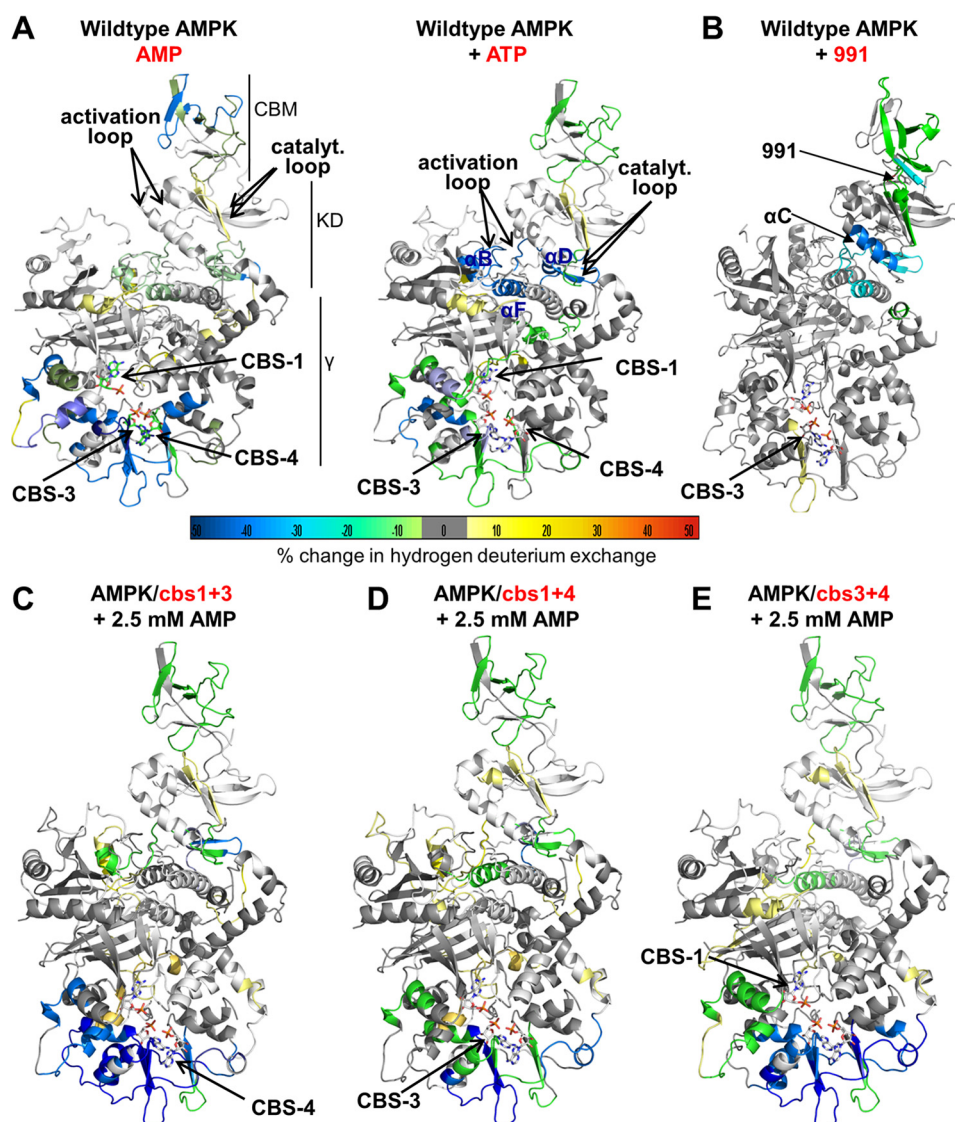


Figure 6. HDX-MS changes of wild-type and mutant AMPK upon incubation with AMP or ATP. Changes were overlaid as heat map onto the structure of $\alpha 1 \beta 2 \gamma 1$ AMPK (4RER). The heat map legend (% change) is shown at the bottom. *A*, AMPK wild-type in the presence versus absence of AMP (left) or ATP (right). *B*, AMPK wild-type in the presence versus absence of 991. *C*, *D*, and *E*, AMPK triple mutants in which only CBS4 (*C*), CBS3 (*D*), or CBS1 (*E*) are functional (see Fig. 1).

conclude that NADPH binds to the high affinity exchangeable AMP-binding site and that this site has a preference for AMP over ATP, suggesting that NADPH binds CBS3.

To physically localize the NADPH-binding site, we collected HDX-MS perturbation data of AMPK in the absence versus presence of 1 mM NADPH. As shown in Fig. 10, NADPH binding selectively protected CBS3 (plus CBS4), but not CBS1, against deuterium exchange. Because CBS4 is non-exchangeable, the HDX-MS data confirm that NADPH specifically binds CBS3. Given the high affinity (low μM IC_{50}) with which AMP competes NADPH from CBS3 and the AMP-over-ATP binding preference of CBS3, we conclude that CBS3, and not as previously proposed CBS1 (19), is the high affinity exchangeable AMP-binding site.

Discussion

AMPK plays critical roles in numerous aspects of physiology (4, 5, 35) and is an important target for the treatment of metabolic diseases, including diabetes and obesity (1, 7), as well as cancer (2, 6, 36) and heart disease (37). In addition, AMPK

activation mimics many of the beneficial effects of exercise and caloric restriction (38–41). Crystal structures and biochemical binding assays have demonstrated that adenine nucleotides can competitively bind to three functional CBS motifs in the γ -subunit, of which CBS3 functions as an adenine sensor site. However, due to the technical difficulties of deconvoluting complex binding events, many aspects of AXP occupancy at individual CBS sites, and how binding to one site is communicated to the other sites, have remained unclear.

To deconvolute the binding events, we generated a set of mutant AMPK complexes in which only individual sites or pairs of sites remain functional, and quantitatively determined their nucleotide-binding and deuterium-exchange patterns. These experiments provided insight into functional and conformational coupling of CBS sites, occupancy in the presence of AXP mixtures, assignment of the high affinity exchangeable AXP site, and a mechanistic model for the requirement of CBS4 for AMP sensing at CBS3.

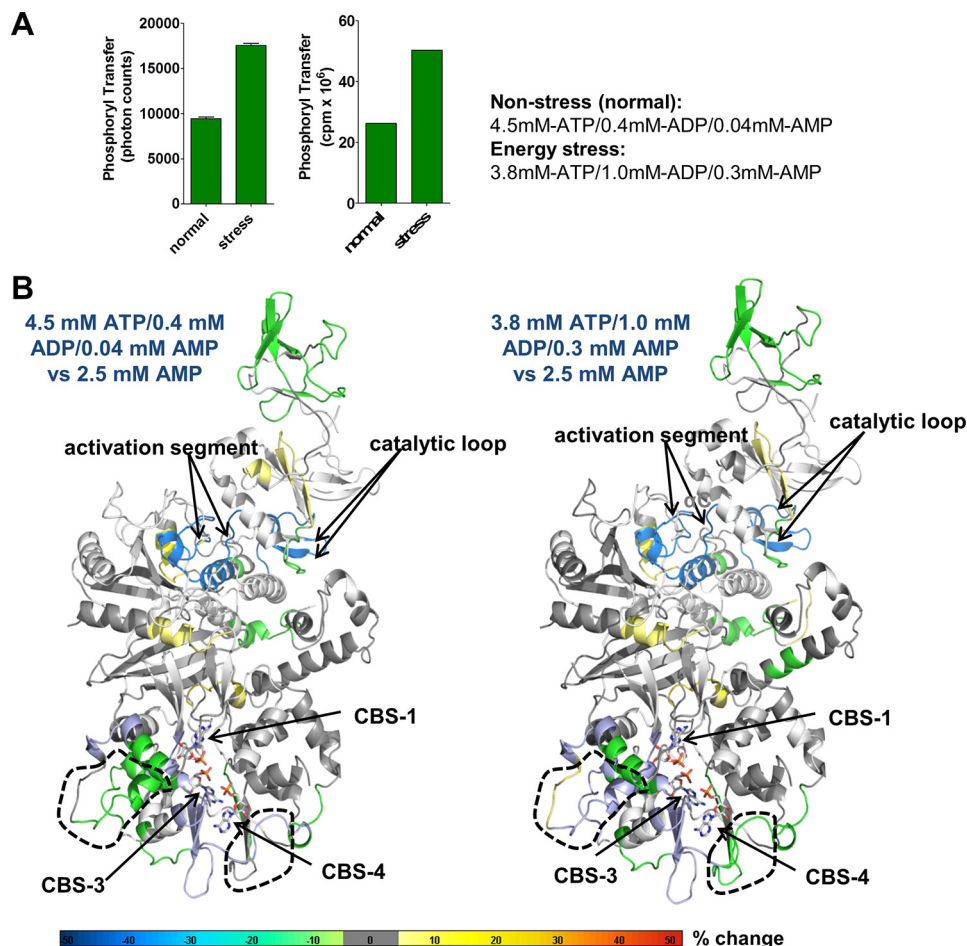


Figure 7. Changes in HDX-MS protection in the presence of AXP mixtures mimicking non-stress and energy stress conditions. A, AMPK kinase assay in the presence of AXP mixtures mimicking nonstress (4.5 mM ATP, 0.4 mM ADP, 0.04 mM AMP) and energy stress (3.8 mM ATP, 1.0 mM ADP, 0.3 mM AMP) conditions. Left panel, AlphaScreen-based kinase assay; right panel, radioactive kinase assay. B, HDX-MS perturbation map of AMPK in the presence of the two different AXP mixtures. HDX-MS changes were overlaid as heat map onto the structure of $\alpha 1\beta 2\gamma 1$ AMPK (PDB code 4RER). Main areas of differential protection are highlighted by dashed outlines. Note that there is no change at the catalytic site, indicating that the catalytic site is constitutively bound by ATP as expected. The color code (% change) is shown below the structures. The relatively mild changes are consistent with noticeable ATP exchange only at CBS3, whereas the non-physiological shift from high concentration of pure AMP to high concentration of pure ATP resulted in strong HDX changes (Fig. 6A), consistent with near full nucleotide exchange at both CBS3, CBS1, and ATP binding/release at the catalytic site.

Functional and conformational coupling of CBS sites

Early binding studies using GST-CBS fusion proteins provided the first support for cooperative AXP binding (18). Here we specifically demonstrate that CBS4 becomes readily exchangeable in the absence of functional CBS1 and CBS3, and AXP-binding to either CBS1 or CBS3 strongly decreased deuterium exchange at CBS4. Therefore, AXP-bound CBS1 and CBS3 can each separately increase AMP affinity at CBS4 and conformationally stabilize CBS4. Consistently, AMPK with mutation of only CBS1 or only CBS3 stably co-purifies with AMP (3) and AXP-binding follows a single site binding model with kinetics different from that of CBS4-only. Stabilization is likely due to His¹⁵¹, which forms hydrogen bonds with the AMP phosphate groups at both CBS4 and CBS1, and His²⁹⁸, which forms hydrogen bonds with the AMP phosphate groups at both CBS4 and CBS3 (Fig. 1). Conversely, CBS3 becomes conformationally stabilized by CBS4 and CBS1, and AMP-binding to CBS4 (functional CBS3 + CBS4 *versus* functional CBS3) increased the CBS3 affinity for AMP by 2 orders of magnitude and increased the AMP/ATP binding preference by 2–3 orders of magnitude.

AMP-sensing requires CBS4 to conformationally stabilize AMP-bound CBS3

Mutating a key adenine-binding residue of CBS4 (I312D) abolishes direct allosteric AMPK activation by AMP (20). This indicates that CBS4 critically contributes to AMP-sensing at CBS3/ α RIM, even though CBS4 is constitutively bound by AMP and does not directly interact with either the α RIM loop or AMP at CBS3. Our fluorescence competition and HDX-MS data suggest that CBS4 is required for AMP sensing by stabilizing AMP binding at CBS3, which increases both the affinity for AMP and the preferential binding of AMP over ATP (Fig. 5). AMP-bound CBS4 therefore allows AMP exchange at CBS3 at nucleotide concentrations that reflect cellular conditions of energy excess ($[ATP]/[AMP] > 100$) and energy stress ($[ATP]/[AMP] \sim 10$ – 20). Our finding that AMP-bound CBS4 selectively stabilizes AMP binding at CBS3 provides a rationale for the CBS4 requirement for AMP sensing.

How does AMP binding to CBS4 mechanistically increase AMP affinity and AMP/ATP-binding preference at CBS3? Inspection of AMPK crystal structures indicates that stabilization is predominantly due to CBS4 positioning the side chain of

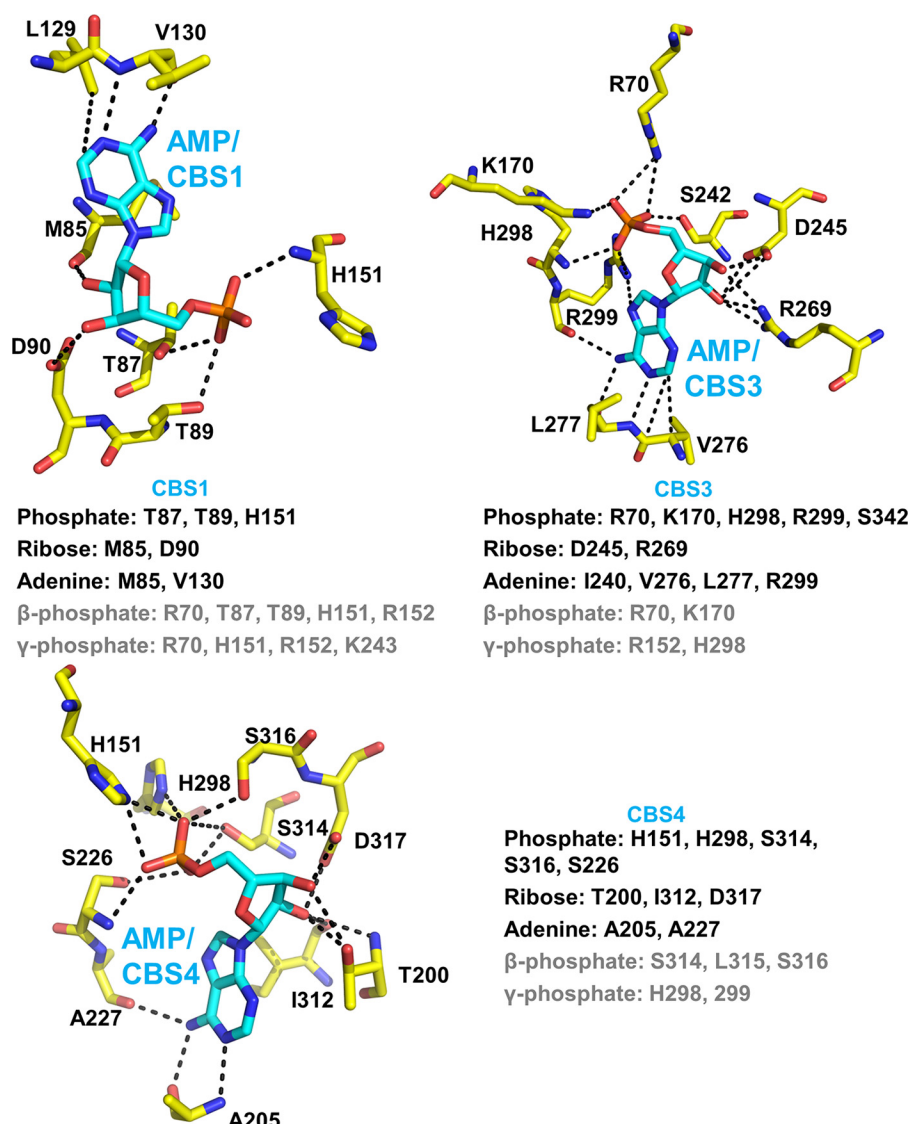


Figure 8. AXP/CBS interaction network in human AMPK-γ1. Shown are the three AMP-bound CBS sites in holo-AMPK co-crystallized with AMP. β- and γ-phosphate interacting residues (gray) are collectively based on the structures of core AMPK co-crystallized with ATP (PDB code 4EAG) (20) and core-AMPK soaked with ATP (PDB codes 2V92 and 4EAG) (19, 20).

His²⁹⁸. The His²⁹⁸ backbone amide directly forms a hydrogen bond with the phosphate group of AMP at CBS3 and positions the adjacent key residue Arg²⁹⁹, which forms two hydrogen bonds with the AMP phosphate group and two with the AMP adenine ring at CBS3 (Fig. 11). Similarly, αRIM Glu³⁶⁴ positions CBS3 Arg⁷⁰ and Lys¹⁷⁰, which form salt bridges with the phosphate group of AMP at CBS3. The phosphate group of AMP carries two negative charges to interact with the three positively charged residues stabilized by αRIM (Arg⁷⁰ and Lys¹⁷⁰) and CBS4 (Arg²⁹⁹) plus the partially positive backbone amide of His²⁹⁸. In contrast, the corresponding α-phosphate of ATP carries only a single charge and therefore interacts less tightly with the positive charge cluster (Fig. 11A). In addition, as seen when structurally aligned with ATP-soaked core-AMPK (PDB code 2V92), the side chains of Arg⁷⁰ and Lys¹⁷⁰ need to move away in opposite directions from Glu³⁶⁴ to avoid clashes with the β- and γ-phosphate groups of ATP (Fig. 11B), thereby weakening the Glu³⁶⁴ interaction. Together, these constraints provide a struc-

tural explanation for both CBS4 and αRIM Glu³⁶⁴ collectively stabilizing AMP preferentially over ATP at CBS3. Consistent with a requirement of both CBS4 and the αRIM for high affinity binding of AMP at CBS3, AMP was preferentially bound to CBS1 and CBS4 in the crystal structure of the αRIM-lacking core-AMPK (19), but preferentially bound to CBS3 and CBS4 in the context of the αRIM-containing holo-AMPK (15, 42).

CBS3 is the high affinity exchangeable AMP-binding site

Our work presents several lines of evidence in support of CBS3 being the high affinity exchangeable AMP-binding site. First, AMP competes NADPH binding with high affinity, whereas ATP competes with moderate affinity, directly indicating that the NADPH-binding site is the high affinity site. This site is CBS3, as NADPH selectively protects CBS3, but not CBS1, against hydrogen deuterium exchange. Second, CBS3 in the presence of functional CBS4 and αRIM binds AMP with high, and ATP with moderate affinity, whereas CBS1 in the

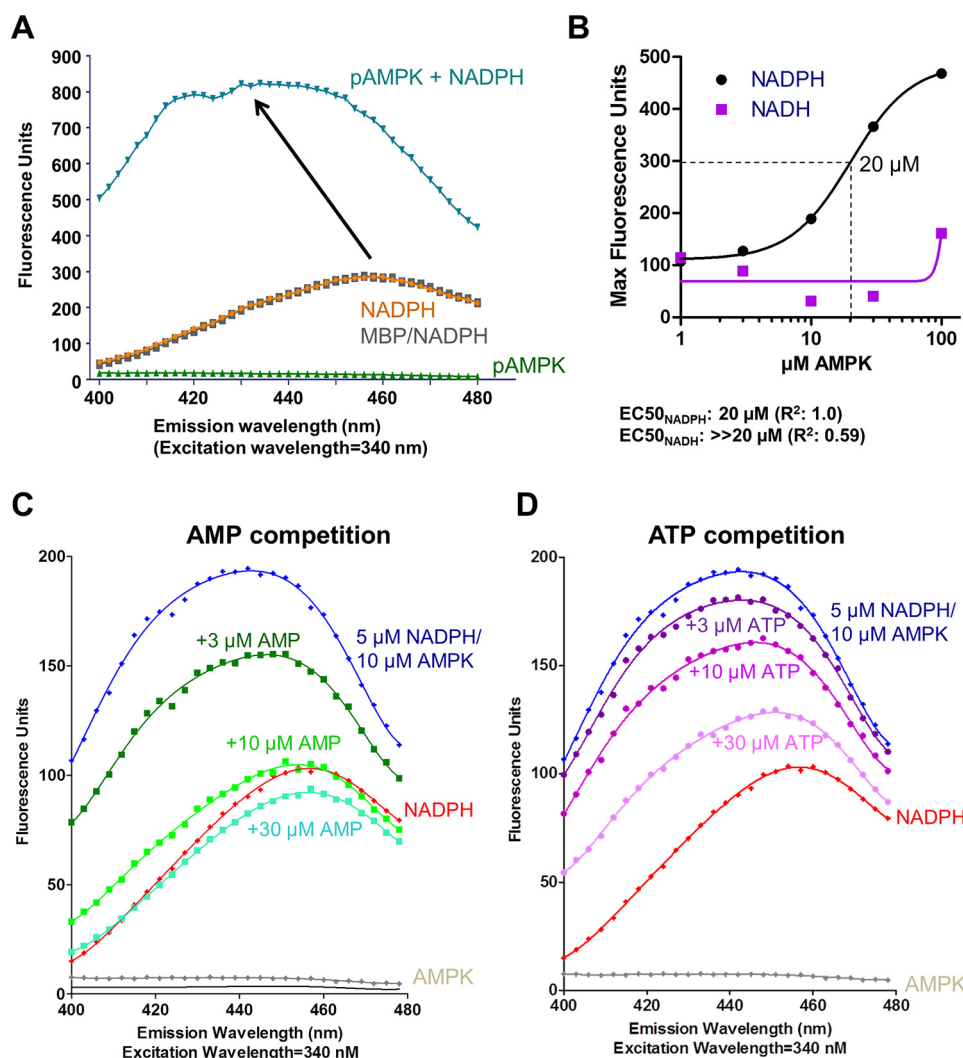


Figure 9. NADPH binds adenine nucleotide-binding site(s) in the γ -subunit. A, NADPH emission spectra in the absence and presence of MBP and MBP-tagged phosphorylated $\alpha 1\beta 2\gamma 1$ AMPK. The spectrum of AMPK in the absence is shown as negative control. B, estimation of NADPH and NADH binding constants. 5 μ M NADPH or NADH were incubated with increasing concentrations of MBP-AMPK and MBP-AMPK concentrations were plotted against fluorescence intensity at the emission maxima. C and D, AMP and ATP compete the NADPH binding signal. Emission spectra of 5 μ M NADPH, 10 μ M AMPK were recorded in the absence and presence of increasing concentrations of AMP (C) and ATP (D).

presence of functional CBS4 binds AMP weakly and with preference of ATP over AMP binding. Third, HDX-MS in the presence of wild-type holo-AMPK demonstrates that AMP stabilizes CBS3 more strongly than ATP, whereas it stabilizes CBS1 less strongly than ATP, consistent with higher affinity AMP binding at CBS3 and relatively stronger ATP binding at CBS1.

Under physiological conditions nucleotide exchange is likely limited to CBS3

Several lines of evidence suggest that during the shift from non-stress (4.5 mM ATP, 0.4 mM ADP, 0.04 mM AMP) to energy stress (3.8 mM ATP, 1.0 mM ADP, 0.3 mM AMP) only CBS3 noticeably exchanges ATP against AMP. By mutating CBS1, we could show that in the presence of the tightly AMP-bound CBS4, CBS3 binds AMP with 10- to 100-fold higher affinity than ATP, closely matching the 10- to 100-fold higher ATP to AMP levels under different stress conditions. In contrast, under the converse conditions, CBS1 binds ATP with similar or higher affinity than AMP, suggesting that at millimolar ATP

and micromolar AMP physiological levels CBS1 remains largely ATP-bound and CBS4 AMP-bound. These conclusions are supported by HDX-MS. First, the AMP/ATP preferences of CBS3 and CBS1 are reflected by CBS3 being more strongly protected by AMP than ATP and, conversely, CBS1 is more strongly protected by ATP than AMP. Second, the AMPK HDX profiles in the sole presence of AMP *versus* ATP differ strongly, consistent with AMP/ATP exchange at both CBS1 and CBS3 as seen in crystal structures and competition assays, whereas the profiles in the presence of stress *versus* non-stress AXP levels are much smaller, indicating reduced AMP/ATP exchange.

ATP is the essential cofactor of numerous cellular reactions, which require ATP concentrations within a very tight range. Yet ATP is constantly and rapidly metabolized (humans hydrolyze roughly their own body weight of ATP per day (43)), and therefore requires highly efficient cellular and organismal regulation of ATP homeostasis. This task is centrally performed by AMPK and depends on the ability of AMPK to precisely and sensitively detect AXP ratios under different physiological con-

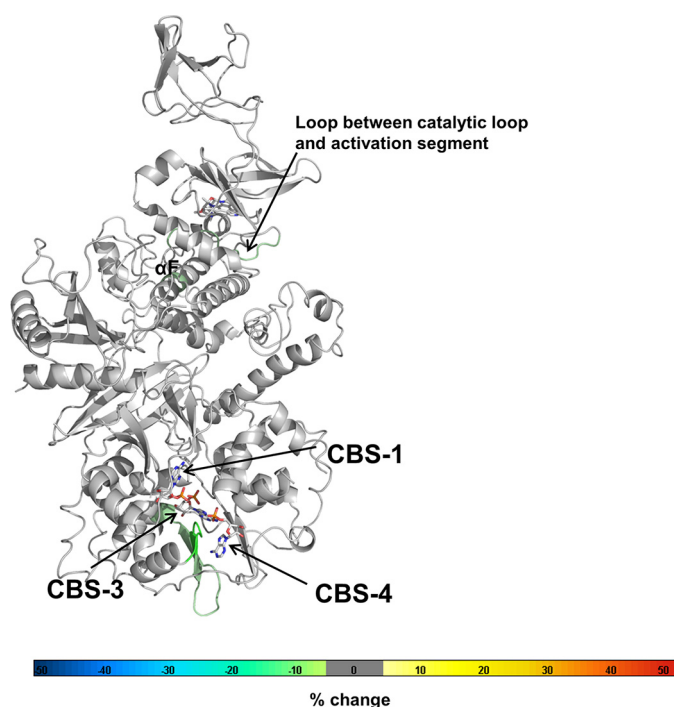


Figure 10. Binding of NADPH protects CBS3 and CBS4. HDX-MS changes wild-type AMPK upon incubation with 1 mM NADPH. Changes were overlaid as heat map onto the structure of $\alpha_1\beta_2\gamma_1$ AMPK (PDB code 4RER). The heat map legend (% change) is shown at the bottom.

ditions to modulate its kinase activity. Our study, together with previous binding assays and crystal structures of holo-AMPK bound to AMP and core-AMPK bound to ATP, provides a first view of the intricate arrangement and interactions between nucleotide-binding sites to allow precise and highly sensitive sensing of AXP ratios under physiological stress and non-stress conditions (Fig. 12). Understanding how the sensed nucleotides modulate kinase activity and activation loop accessibility will require the missing key structure of holo-AMPK in its inactive ATP-bound state.

Experimental procedures

DNA constructs and reagents

The human wild-type His₆- $\alpha_1(11-550)\beta_2\gamma_1$ AMPK expression construct was described previously (32). Mutations were introduced by site-directed mutagenesis using the QuikChange method (Stratagene) or standard PCR-based methods. All expression constructs and mutations were confirmed by DNA sequencing.

Protein expression and purification

Expression plasmids were transformed into *Escherichia coli* BL21(DE3). Cells were grown to an A_{600} of ~ 1 at 28 °C and induced with 100 μ M isopropyl β -D-thio-galactopyranoside at 16 °C overnight. Cell pellets were resuspended in 25 mM Tris, pH 8.0, 300 mM NaCl, 25 mM imidazole, 10% glycerol, 5 mM β -mercaptoethanol, and lysed by French Press with pressure set to 900 pascal. Lysates were cleared by centrifugation for 30 min at 20,000 $\times g$, passed over a 50-ml nickel-chelating HP-Sepharose column (GE Healthcare), and eluted with 25 mM Tris, pH 8.0, 300 mM NaCl, 500 mM imidazole, 10% glycerol, 5

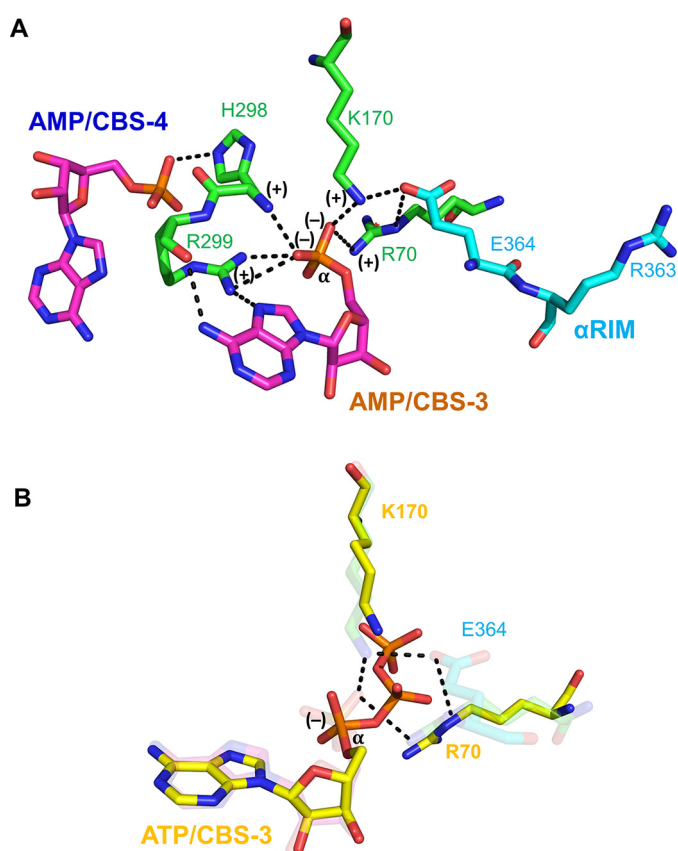


Figure 11. CBS4 and α RIM stabilize AMP-binding at CBS3 and increase the AMP/ATP-binding preference. A, CBS4-CBS3- α RIM network in AMP-bound AMPK (PDB code 4CFE). B, transparent structure from panel A overlaid with ATP, Arg⁷⁰, and Lys¹⁷⁰ from the structure of ATP-soaked core-AMPK (PDB code 2V92). Note that the α -phosphate of ATP has only a single negative charge and that the β - and γ -phosphate groups of ATP push Lys¹⁷⁰ and Arg⁷⁰ away from α RIM Glu³⁶⁴.

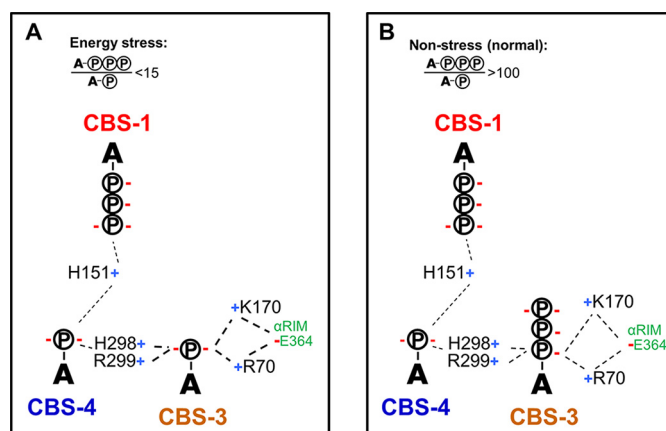


Figure 12. Schematic model of AXP occupancy under energy-stress (A) and non-stress (B) conditions. Positive and negative charges are indicated by blue "+" and red "-" signs. A-circled P, AMP; A-3 circled P, ATP.

mM β -mercaptoethanol. The eluted His-tagged AMPK was further purified by size-exclusion chromatography through a 300-ml HiLoad 26/60 Superdex 200 column (GE Healthcare) in 10 mM Tris, pH 8.0, 150 mM NaCl, 5 mM MgCl₂, 1 mM EDTA, 10% glycerol, 2 mM DTT. The proteins eluted from the gel filtration column at a volume corresponding to the size of a monomeric complex at a purity $\geq 95\%$ as judged by SDS-PAGE (Fig. 2).

Deconvoluting AMPK adenine nucleotide binding and sensing

Fluorescence binding and competition assays

Deac-ADP was synthesized by the method of Webb *et al.* (44). 4 μM AMPK wild-type and mutant proteins were incubated with 0.5 μM deac-ADP in 10 mM Tris, pH 8.0, 150 mM NaCl, 1 mM EDTA, 10% glycerol, 2 mM DTT in the presence and absence of 5 mM MgCl_2 for 30 min at room temperature. The emission spectrum of deac-ADP was recorded using a SpectraMax M2^e reader (Molecular Devices Corporation). For deac-ADP competition assays, AMP or ATP were added at increasing concentrations, whereas deac-ADP was kept at a constant concentration (0.5 μM). The IC_{50} values were obtained from curve fitting to the competitive inhibitor model using GraphPad Prism.

For NADPH emission spectra, 5 μM NADPH were incubated with either 10 μM MBP or 10 μM MBP-AMPK and excited at 340 nm using the SpectraMax M2^e cuvette mode. For binding isotherms, NADP and NADPH (5 μM each) were incubated with increasing concentrations of AMPK, excited at 340 nm, and full emission spectra were recorded. Fluorescence emission maxima were plotted against AMPK concentration and EC_{50} values were obtained from curve fitting to the one-site binding model using GraphPad Prism.

HDX coupled with mass spectrometry

Solution-phase amide HDX experiments were carried out using a fully automated system as described previously (45, 46). Five microliters of a 10 μM protein solution were mixed with 20 μl of D_2O containing HDX buffer (20 mM KPO_4 , pH 7.4, 50 mM KCl) and incubated at 4 °C for 10, 30, 60, 300, 900, and 3,600 s. Following on exchange, unwanted forward or back exchange was minimized and the protein was denatured by dilution with 25 μl of quench solution (0.1%, v/v, trifluoroacetic acid (TFA) in 3 M urea). Samples were then passed through an immobilized pepsin column (prepared in house) (47) at 200 $\mu\text{l}/\text{min}$ (0.1%, v/v, TFA, 15 °C) and the resulting peptides were trapped on a C8 trap column (Hypersil Gold, Thermo Fisher). The bound peptides were then gradient eluted (4–40%, w/v, CH_3CN and 0.3%, w/v, formic acid) across a 2 \times 50-mm C18 high performance liquid chromatography column (Hypersil Gold, Thermo Fisher) for 5 min at 4 °C. The eluted peptides were then subjected to electrospray ionization directly coupled with a high resolution Orbitrap mass spectrometer (Exactive, Thermo Fisher). Each HDX experiment was carried out in triplicate. Peptide ion signals were confirmed if they had a MASCOT score of 20 or greater and had no ambiguous hits using a decoy (reverse) sequence in a separate experiment using a 60-min gradient. The intensity-weighted average m/z value (centroid) of each isotopic envelope of the peptide was calculated with in house HDX Workbench software (48) and corrected for back-exchange on an estimated 70% recovery and accounting for the known deuterium content of the on-exchange buffer.

Isothermal titration calorimetry (ITC)

ITC was performed in a MicroCal PEAQ iTC200 (Malvern) with 18 injections at room temperature. 30 μM AMPK was loaded into the cell and 300 μM AMP in the syringe. Data analysis was performed with the MicroCal PEAQ-ITC instrument control software and yielded: $n = 0.856 \pm 0.02$ sites, $K_d =$

$2.06 \pm 0.538 \mu\text{M}$, $\Delta H = 12.6 \pm 0.757 \text{ kcal/mol}$, $\Delta G = -7.76$, $-T\Delta S = 4.88 \text{ kcal/mol}$.

Kinase assays

For the radioactive kinase assay, 10 nM phosphorylated AMPK were incubated with 15 μM biotin-SAMS peptide, 2 mM DTT, and 0.25 μl [γ - ^{32}P]ATP per 15- μl reaction in kinase buffer (25 mM Tris, pH 7.4, 12 mM MgCl_2 , 1 mM Na_3VO_4 , 5 mM NaF) in the presence of AXP mixtures mimicking non-stress (4.5 mM ATP, 0.4 mM ADP, 0.04 mM AMP) and energy-stress (3.8 mM ATP, 1.0 mM ADP, 0.3 mM AMP) conditions for 30 min at room temperature. Reactions were terminated by addition of 0.5 volumes of 7.5 M guanidine hydrochloride solution in water and reactions were spotted on a SAM 2[®] Biotin Capture Membrane (Promega). The membrane was washed once for 30 s with 2 M NaCl, 3 times for 2 min with 2 M NaCl, 4 times for 2 min with 2 M NaCl in 1% H_3PO_4 , and 2 times for 30 s with deionized water to remove unbound reaction components. After drying the membrane at room temperature for 30–60 min, signals were quantitated by phosphorimager analysis.

The luminescence-proximity AlphaScreen-based kinase assay will be described in detail in a future publication and is based on a previously described AMPK activity reporter assay (49). Briefly, 10 nM phosphorylated AMPK were incubated with 50 μM of a biotinylated peptide substrate (biotin-GSTKMR-RVATLVLDLGYKK) on ice for 10 min. The reaction was terminated by 1000-fold dilution in AlphaScreen buffer (50 mM MOPS, pH 7.4, 50 mM NaF, 50 mM CHAPS, and 0.1 mg/ml of bovine serum albumin) and further incubated in the presence of 50 nM His₆-Rad53 FHA domain protein, 5 $\mu\text{g}/\text{ml}$ each of AlphaScreen streptavidin donor beads and nickel-acceptor beads at room temperature for 90 min in the presence of AXP mixtures mimicking non-stress (4.5 mM ATP, 0.4 mM ADP, 0.04 mM AMP) or energy-stress (3.8 mM ATP, 1.0 mM ADP, 0.3 mM AMP) conditions. Donor beads contain a photosensitizer, which can convert ambient oxygen into short-lived singlet oxygen upon light activation at 680 nm. When the acceptor beads are brought close enough to the donor beads by interaction between His₆-tagged FHA domain protein and AMPK-phosphorylated biotinylated peptide substrate, singlet oxygen can diffuse from the donor to the acceptor beads and transfer energy to the thioxene derivatives of the acceptor beads, resulting in light emission at 520 to 620 nm, which was measured in an Envision (PerkinElmer Life Sciences) plate reader.

Author contributions—X. G., Y. Y., S. J. N., A. K., D. G., J. K., M. H. E. T., P. W. deW., L. W., and X. L. conducted the experiments, M. R. W., P. R. G., H. E. X., and K. M. analyzed the results, and K. M. wrote the paper with comments from all authors.

Acknowledgments—We thank Stoyan Milev (Malvern Instruments) for help with the ITC experiment and Michelle Martin for administrative support.

References

1. Bullon, P., Marin-Aguilar, F., and Roman-Malo, L. (2016) AMPK/mitochondria in metabolic diseases. *EXS* 107, 129–152

2. Cheng, J., Zhang, T., Ji, H., Tao, K., Guo, J., and Wei, W. (2016) Functional characterization of AMP-activated protein kinase signaling in tumorigenesis. *Biochim. Biophys. Acta* **1866**, 232–251
3. Hardie, D. G., Ross, F. A., and Hawley, S. A. (2012) AMP-activated protein kinase: a target for drugs both ancient and modern. *Chem. Biol.* **19**, 1222–1236
4. Mihaylova, M. M., and Shaw, R. J. (2011) The AMPK signalling pathway coordinates cell growth, autophagy and metabolism. *Nat. Cell. Biol.* **13**, 1016–1023
5. Steinberg, G. R., and Kemp, B. E. (2009) AMPK in health and disease. *Physiol. Rev.* **89**, 1025–1078
6. Wang, Z., Wang, N., Liu, P., and Xie, X. (2016) AMPK and cancer. *EXS* **107**, 203–226
7. Weikel, K. A., Ruderman, N. B., and Cacicedo, J. M. (2016) Unraveling the actions of AMP-activated protein kinase in metabolic diseases: systemic to molecular insights. *Metabolism* **65**, 634–645
8. Carling, D., Clarke, P. R., Zammit, V. A., and Hardie, D. G. (1989) Purification and characterization of the AMP-activated protein kinase: copurification of acetyl-CoA carboxylase kinase and 3-hydroxy-3-methylglutaryl-CoA reductase kinase activities. *Eur. J. Biochem.* **186**, 129–136
9. Cheung, P. C., Salt, I. P., Davies, S. P., Hardie, D. G., and Carling, D. (2000) Characterization of AMP-activated protein kinase γ -subunit isoforms and their role in AMP binding. *Biochem. J.* **346**, 659–669
10. Oakhill, J. S., Chen, Z. P., Scott, J. W., Steel, R., Castelli, L. A., Ling, N., Macaulay, S. L., and Kemp, B. E. (2010) β -Subunit myristoylation is the gatekeeper for initiating metabolic stress sensing by AMP-activated protein kinase (AMPK). *Proc. Natl. Acad. Sci. U.S.A.* **107**, 19237–19241
11. Zhang, Y. L., Guo, H., Zhang, C. S., Lin, S. Y., Yin, Z., Peng, Y., Luo, H., Shi, Y., Lian, G., Zhang, C., Li, M., Ye, Z., Ye, J., Han, J., Li, P., Wu, J. W., and Lin, S. C. (2013) AMP as a low-energy charge signal autonomously initiates assembly of AXIN-AMPK-LKB1 complex for AMPK activation. *Cell Metab.* **18**, 546–555
12. Davies, S. P., Helps, N. R., Cohen, P. T., and Hardie, D. G. (1995) 5'-AMP inhibits dephosphorylation, as well as promoting phosphorylation, of the AMP-activated protein kinase. Studies using bacterially expressed human protein phosphatase-2C α and native bovine protein phosphatase-2AC. *FEBS Lett.* **377**, 421–425
13. Yeh, L. A., Lee, K. H., and Kim, K. H. (1980) Regulation of rat liver acetyl-CoA carboxylase: regulation of phosphorylation and inactivation of acetyl-CoA carboxylase by the adenylate energy charge. *J. Biol. Chem.* **255**, 2308–2314
14. Gowans, G. J., Hawley, S. A., Ross, F. A., and Hardie, D. G. (2013) AMP is a true physiological regulator of AMP-activated protein kinase by both allosteric activation and enhancing net phosphorylation. *Cell Metab.* **18**, 556–566
15. Xiao, B., Sanders, M. J., Underwood, E., Heath, R., Mayer, F. V., Carmena, D., Jing, C., Walker, P. A., Eccleston, J. F., Haire, L. F., Saiu, P., Howell, S. A., Aasland, R., Martin, S. R., Carling, D., and Gamblin, S. J. (2011) Structure of mammalian AMPK and its regulation by ADP. *Nature* **472**, 230–233
16. Ross, F. A., Jensen, T. E., and Hardie, D. G. (2016) Differential regulation by AMP and ADP of AMPK complexes containing different γ subunit isoforms. *Biochem. J.* **473**, 189–199
17. Oakhill, J. S., Steel, R., Chen, Z. P., Scott, J. W., Ling, N., Tam, S., and Kemp, B. E. (2011) AMPK is a direct adenylate charge-regulated protein kinase. *Science* **332**, 1433–1435
18. Scott, J. W., Hawley, S. A., Green, K. A., Anis, M., Stewart, G., Scullion, G. A., Norman, D. G., and Hardie, D. G. (2004) CBS domains form energy-sensing modules whose binding of adenosine ligands is disrupted by disease mutations. *J. Clin. Invest.* **113**, 274–284
19. Xiao, B., Heath, R., Saiu, P., Leiper, F. C., Leone, P., Jing, C., Walker, P. A., Haire, L., Eccleston, J. F., Davis, C. T., Martin, S. R., Carling, D., and Gamblin, S. J. (2007) Structural basis for AMP binding to mammalian AMP-activated protein kinase. *Nature* **449**, 496–500
20. Chen, L., Wang, J., Zhang, Y. Y., Yan, S. F., Neumann, D., Schlattner, U., Wang, Z. X., and Wu, J. W. (2012) AMP-activated protein kinase undergoes nucleotide-dependent conformational changes. *Nat. Struct. Mol. Biol.* **19**, 716–718
21. Calabrese, M. F., Rajamohan, F., Harris, M. S., Caspers, N. L., Magyar, R., Withka, J. M., Wang, H., Borzilleri, K. A., Sahasrabudhe, P. V., Hoth, L. R., Geoghegan, K. F., Han, S., Brown, J., Subashi, T. A., Reyes, A. R., et al. (2014) Structural basis for AMPK activation: natural and synthetic ligands regulate kinase activity from opposite poles by different molecular mechanisms. *Structure* **22**, 1161–1172
22. Rajamohan, F., Reyes, A. R., Frisbie, R. K., Hoth, L. R., Sahasrabudhe, P., Magyar, R., Landro, J. A., Withka, J. M., Caspers, N. L., Calabrese, M. F., Ward, J., and Kurumbail, R. G. (2016) Probing the enzyme kinetics, allosteric modulation and activation of α 1- and α 2-subunit-containing AMP-activated protein kinase (AMPK) heterotrimeric complexes by pharmacological and physiological activators. *Biochem. J.* **473**, 581–592
23. Arad, M., Benson, D. W., Perez-Atayde, A. R., McKenna, W. J., Sparks, E. A., Kanter, R. J., McGarry, K., Seidman, J. G., and Seidman, C. E. (2002) Constitutively active AMP kinase mutations cause glycogen storage disease mimicking hypertrophic cardiomyopathy. *J. Clin. Invest.* **109**, 357–362
24. Daniel, T., and Carling, D. (2002) Functional analysis of mutations in the γ 2 subunit of AMP-activated protein kinase associated with cardiac hypertrophy and Wolff-Parkinson-White syndrome. *J. Biol. Chem.* **277**, 51017–51024
25. Hinson, J. T., Chopra, A., Lowe, A., Sheng, C. C., Gupta, R. M., Kuppusamy, R., O'Sullivan, J., Rowe, G., Wakimoto, H., Gorham, J., Zhang, K., Musunuru, K., Gerszten, R. E., Wu, S. M., Chen, C. S., et al. (2016) Integrative analysis of PRKAG2 cardiomyopathy iPSC and microtissue models identifies AMPK as a regulator of metabolism, survival, and fibrosis. *Cell Rep.* **17**, 3292–3304
26. Kim, M., Hunter, R. W., Garcia-Menendez, L., Gong, G., Yang, Y. Y., Kolwicz, S. C., Jr., Xu, J., Sakamoto, K., Wang, W., and Tian, R. (2014) Mutation in the γ 2-subunit of AMP-activated protein kinase stimulates cardiomyocyte proliferation and hypertrophy independent of glycogen storage. *Circ. Res.* **114**, 966–975
27. Yang, X., Mudgett, J., Bou-About, G., Champy, M. F., Jacobs, H., Monnassier, L., Pavlovic, G., Sorg, T., Herault, Y., Petit-Demoulière, B., Lu, K., Feng, W., Wang, H., Ma, L. J., Askew, R., et al. (2016) Physiological expression of AMPK γ 2RG mutation causes Wolff-Parkinson-White syndrome and induces kidney injury in mice. *J. Biol. Chem.* **291**, 23428–23439
28. Granlund, A., Kotova, O., Benziene, B., Galuska, D., Jensen-Waern, M., Chibalin, A. V., and Essén-Gustavsson, B. (2010) Effects of exercise on muscle glycogen synthesis signalling and enzyme activities in pigs carrying the PRKAG3 mutation. *Exp. Physiol.* **95**, 541–549
29. Costford, S. R., Kavaslar, N., Ahituv, N., Chaudhry, S. N., Schackwitz, W. S., Dent, R., Pennacchio, L. A., McPherson, R., and Harper, M. E. (2007) Gain-of-function R225W mutation in human AMPK γ 3 causing increased glycogen and decreased triglyceride in skeletal muscle. *PLoS ONE* **2**, e903
30. Crawford, S. A., Costford, S. R., Aguer, C., Thomas, S. C., deKemp, R. A., DaSilva, J. N., Lafontaine, D., Kendall, M., Dent, R., Beanlands, R. S., McPherson, R., and Harper, M. E. (2010) Naturally occurring R225W mutation of the gene encoding AMPK-activated protein kinase (AMPK) γ 3 results in increased oxidative capacity and glucose uptake in human primary myotubes. *Diabetologia* **53**, 1986–1997
31. Chen, L., Xin, F. J., Wang, J., Hu, J., Zhang, Y. Y., Wan, S., Cao, L. S., Lu, C., Li, P., Yan, S. F., Neumann, D., Schlattner, U., Xia, B., Wang, Z. X., and Wu, J. W. (2013) Conserved regulatory elements in AMPK. *Nature* **498**, E8–E10
32. Li, X., Wang, L., Zhou, X. E., Ke, J., de Waal, P. W., Gu, X., Tan, M. H., Wang, D., Wu, D., Xu, H. E., and Melcher, K. (2015) Structural basis of AMPK regulation by adenine nucleotides and glycogen. *Cell Res.* **25**, 50–66
33. Forgacs, E., Cartwright, S., Kovács, M., Sakamoto, T., Sellers, J. R., Corrie, J. E., Webb, M. R., and White, H. D. (2006) Kinetic mechanism of myosinV-S1 using a new fluorescent ATP analogue. *Biochemistry* **45**, 13035–13045
34. Xiao, B., Sanders, M. J., Carmena, D., Bright, N. J., Haire, L. F., Underwood, E., Patel, B. R., Heath, R. B., Walker, P. A., Hallen, S., Giordanetto, F., Martin, S. R., Carling, D., and Gamblin, S. J. (2013) Structural basis of AMPK regulation by small molecule activators. *Nat. Commun.* **4**, 3017
35. Hardie, D. G. (2011) AMP-activated protein kinase: an energy sensor that regulates all aspects of cell function. *Genes Dev.* **25**, 1895–1908

36. Faubert, B., Vincent, E. E., Poffenberger, M. C., and Jones, R. G. (2015) The AMP-activated protein kinase (AMPK) and cancer: many faces of a metabolic regulator. *Cancer Lett.* **356**, 165–170
37. Daskalopoulos, E. P., Dufey, C., Beauloye, C., Bertrand, L., and Horman, S. (2016) AMPK in cardiovascular diseases. *EXS* **107**, 179–201
38. Wall, C. E., Yu, R. T., Atkins, A. R., Downes, M., and Evans, R. M. (2016) Nuclear receptors and AMPK: can exercise mimetics cure diabetes? *J. Mol. Endocrinol.* **57**, R49–R58
39. Burkewitz, K., Morantte, I., Weir, H. J., Yeo, R., Zhang, Y., Huynh, F. K., Ilkayeva, O. R., Hirschey, M. D., Grant, A. R., and Mair, W. B. (2015) Neuronal CRTC-1 governs systemic mitochondrial metabolism and lifespan via a catecholamine signal. *Cell* **160**, 842–855
40. Mair, W., Morantte, I., Rodrigues, A. P., Manning, G., Montminy, M., Shaw, R. J., and Dillin, A. (2011) Lifespan extension induced by AMPK and calcineurin is mediated by CRTC-1 and CREB. *Nature* **470**, 404–408
41. Narkar, V. A., Downes, M., Yu, R. T., Embler, E., Wang, Y. X., Banayo, E., Mihaylova, M. M., Nelson, M. C., Zou, Y., Juguilon, H., Kang, H., Shaw, R. J., and Evans, R. M. (2008) AMPK and PPAR δ agonists are exercise mimetics. *Cell* **134**, 405–415
42. Langendorf, C. G., Ngoei, K. R., Scott, J. W., Ling, N. X., Issa, S. M., Gorman, M. A., Parker, M. W., Sakamoto, K., Oakhill, J. S., and Kemp, B. E. (2016) Structural basis of allosteric and synergistic activation of AMPK by furan-2-phosphonic derivative C2 binding. *Nat. Commun.* **7**, 10912
43. Di Carlo, S. E., and Collins, H. L. (2001) Submitting illuminations for review. *Adv. Physiol. Educ.* **25**, 70–71
44. Webb, M. R., Reid, G. P., Munasinghe, V. R., and Corrie, J. E. (2004) A series of related nucleotide analogues that aids optimization of fluorescence signals in probing the mechanism of P-loop ATPases, such as actomyosin. *Biochemistry* **43**, 14463–14471
45. Chalmers, M. J., Busby, S. A., Pascal, B. D., He, Y., Hendrickson, C. L., Marshall, A. G., and Griffin, P. R. (2006) Probing protein ligand interactions by automated hydrogen/deuterium exchange mass spectrometry. *Anal. Chem.* **78**, 1005–1014
46. Goswami, D., Devarakonda, S., Chalmers, M. J., Pascal, B. D., Spiegelman, B. M., and Griffin, P. R. (2013) Time window expansion for HDX analysis of an intrinsically disordered protein. *J. Am. Soc. Mass Spectrom.* **24**, 1584–1592
47. Chalmers, M. J., Busby, S. A., Pascal, B. D., Southern, M. R., and Griffin, P. R. (2007) A two-stage differential hydrogen deuterium exchange method for the rapid characterization of protein/ligand interactions. *J. Biomol. Tech.* **18**, 194–204
48. Pascal, B. D., Willis, S., Lauer, J. L., Landgraf, R. R., West, G. M., Marciano, D., Novick, S., Goswami, D., Chalmers, M. J., and Griffin, P. R. (2012) HDX workbench: software for the analysis of H/D exchange MS data. *J. Am. Soc. Mass Spectrom.* **23**, 1512–1521
49. Tsou, P., Zheng, B., Hsu, C. H., Sasaki, A. T., and Cantley, L. C. (2011) A fluorescent reporter of AMPK activity and cellular energy stress. *Cell Metab.* **13**, 476–486

Deconvoluting AMP-activated protein kinase (AMPK) adenine nucleotide binding and sensing

Xin Gu, Yan Yan, Scott J. Novick, Amanda Kovach, Devrishi Goswami, Jiyuan Ke, M. H. Eileen Tan, Lili Wang, Xiaodan Li, Parker W. de Waal, Martin R. Webb, Patrick R. Griffin, H. Eric Xu and Karsten Melcher

J. Biol. Chem. 2017, 292:12653-12666.

doi: 10.1074/jbc.M117.793018 originally published online June 14, 2017

Access the most updated version of this article at doi: [10.1074/jbc.M117.793018](https://doi.org/10.1074/jbc.M117.793018)

Alerts:

- [When this article is cited](#)
- [When a correction for this article is posted](#)

[Click here](#) to choose from all of JBC's e-mail alerts

Supplemental material:

<http://www.jbc.org/content/suppl/2017/06/14/M117.793018.DC1>

This article cites 49 references, 11 of which can be accessed free at <http://www.jbc.org/content/292/30/12653.full.html#ref-list-1>